



DIAGNOSTIC AUTOMATION, INC.

23961 Craftsman Road, Suite E/F, Calabasas, CA 91302

Tel: (818) 591-3030 Fax: (818) 591-8383

onestep@rapidtest.com

technicalsupport@rapidtest.com

www.rapidtest.com

IVD



See external label



2°C-8°C



Σ=96 tests

REF

#2032-14

OLAB

Enzyme immunoassay for the quantitative
determination of human autoantibodies against oxidized
low-density lipoprotein in serum

Cat. No.2032-14

1. Introduction

Oxidized low density lipoprotein (oLDL) is believed to play a critical role in the development and progression of atherosclerosis (1-3). Accumulation of oLDL in macrophages and smooth muscle cells causes foam cell formation, an initial step in the disease (4-6). Recent evidence suggests that autoantibodies against oxidatively modified LDL can be used as a parameter that consistently mirrors the occurrence of oxidation processes taking place in vivo. In fact, elevated levels of autoantibodies against oLDL have been detected in the blood stream of patients with coronary artery disease (7). Moreover, recent studies indicate a correlation between autoantibodies against oLDL and the progression of carotid atherosclerosis (8). Increased serum concentrations of oLAB have also been described in various diseases such as pre-eclampsia and systemic lupus erythematosus (9, 10). Decreased oLAB titers were observed during septicemia and myocardial infarction (11, 12).

An overview on the clinical applications of oLAB has recently been published (11).

2. Principle of the assay

The oLAB kit is an enzyme immunoassay designed to determine human autoantibodies to oxidized LDL directly in serum. Cu ++ oxidized LDL is coated onto microtiter strips as antigen (13, 14). Autoantibodies, if present in the prediluted serum, bind specifically to the antigen. After a washing step, a specific peroxidase conjugated anti human IgG antibody detects the presence of bound autoantibodies. After removal of unbound conjugate through washing, tetramethylbenzidine (TMB) is added to the wells as a non toxic chromogenic substrate. The concentration of specific IgG in the sample is quantitated by an enzyme catalyzed color change detectable on a standard ELISA reader. The amount of color developed is directly proportional to the concentration of antibodies in the sample. The assay is standardized with defined amounts of oLAB in a serum matrix. The IgG concentration in the samples is quantitated in milliunits (mU), based on a characterized serum with a high oLAB titer and a low cross reactivity with native LDL (< 5%).

3. Contents of the kit

Store all reagents refrigerated at 4 °C. Refer to the expiry date on the kit box.

1 uncoated microwell plate for sample dilution

2 x 6 microwell strips
each 8 well strip is coated with oxidized LDL; packed in alubag with desiccant; 1 stripholder

Assay buffer
The vial contains 120 ml assay buffer ready for use

Washing buffer
The vial contains 100 ml of 10 fold buffer concentrate

Conjugate

The vial contains 12 ml mouse monoclonal anti-human IgG fc specific horseradish peroxidase conjugate, ready for use

6 standards

The vials contain 1200, 600, 300, 150, 75, 37 mU/ml oLAB in 0,5 ml human serum, ready for use

2 controls

The vials contain about 1000 and 300 mU/ml oLAB in 0,5 ml human serum, ready for use

Substrate

The vial contains 12 ml TMB solution, ready for use

Stop solution

The vial contains 7 ml of stop solution, ready for use (diluted sulphuric acid)

2 self-adhesive plastic films

1 protocol sheet

1 package insert (instructions for use)

4. Additional material and equipment required

Distilled water

Measuring cylinder

Pipettes for 200 μ l and 50 μ l with disposable tips

Multichannel pipette or multipette for 20 μ l, 100 μ l and 200 μ l

Incubator for 37 °C

Manual or automatic microwell washer

ELISA reader equipped with 450 nm filter

Semilogarithmic paper or software for calculation of results

5. Sample preparation

Store freshly collected serum samples at -20 °C if not assayed on the same day. Do not use lipemic or hemolytic samples. This test system is not designed for plasma.

Preparation of reagents:

Allow all reagents to reach room temperature (18 - 26 °C) before use

Dilute washing buffer concentrate 1:10 (1 part buffer + 9 parts distilled water) to a final volume of 1000 ml with distilled water. Diluted buffer is stable at 4 °C until expiry date stated on the label.

Store coated strips in a desiccator after removal from the foil pouch. Strips will be stable at room temperature (18-26 °C) until expiry date stated on the label.

6. Performance of the assay

We recommend duplicates for all values. Standards, controls and samples must be prediluted 1:50 according to the following protocol:

Mark positions for blank, standards, controls and samples on the protocol sheet supplied

Take microtiter strips out of the alubag (plate 1) and mark as appropriate. Mark two wells as blank

Mark uncoated microwell plate exactly in the same way (plate 2 - for predilution)

Pipette 200 µl of assay buffer into all marked wells of the coated microwell plate (plate 1) incl. blank

Pipette 200 µl of assay buffer into all marked wells of the uncoated microwell plate (plate 2)

Pipette 50 µl of standard, control or sample in respective wells of plate 2

Shake well

Transfer 20 µl of the prediluted samples to the coated strips (Plate 1) with a multichannel pipette
THIS STEP MUST BE COMPLETED WITHIN 15 MINUTES!

Cover strips with plastic film and incubate for 90 minutes at 37 °C

Discard contents of the wells and wash 4 x with 300 µl diluted washing buffer

Add 100 µl conjugate to all wells including blank

Cover strips with plastic film and incubate for
30 minutes at room temperature

Discard contents of the wells and wash 4 x with 300 µl diluted buffer

Add 100 µl substrate to all wells including blank

Incubate strips for 15 minutes at room temperature in the dark

Add 50 µl stop solution to all wells

Shake well and determine absorption with an ELISA reader at 450 nm against 690 or against 620 nm as reference

7. Calculation of results

The extinction of the blank is subtracted from all other values. A calibration curve is constructed from the standards. Commercially available software can be used as well as semilogarithmic paper. Results of the samples are read from this calibration curve. If the concentration of oLAB in the sample exceeds 1100 mU/ml, further dilution and measurement of the diluted sample is recommended.

8. Assay characteristics

Standard range: 37 - 1200 mU/ml

Sample: 50 µl serum

Assay time: 3 hours

Storage: 4 °C

Precision: Intraassay

Mean value (mU/ml)	CV	Number
119 ± 4	3,6%	n = 8
324 ± 14	4,3%	n = 8

Interassay

Mean value (mU/ml)	CV	Number
139 ± 11	8,2%	n = 5
544 ± 22	4,0%	n = 5

9. Precautions

All test components of human source were tested with 3rd generation tests against HIV-Ab and HBsAG; all components were found to be negative. However, standards as well as controls should be handled and disposed as if they were infectious, since no test method can offer complete assurance.

Do not interchange kit components from different lots

Do not use kit components beyond the expiry date

Protect reagents from direct sunlight

Do not pipette by mouth

Do not eat, drink, smoke or apply cosmetics where reagents are used

Avoid all contact with the reagents by using gloves

Sulphuric acid is irritating to eyes and skin. Flush with water if contact occurs

All liquid reagents contain 0,01% thimerosal as preservative. Avoid contact with skin and mucous membrane.

10. References

1. Steinberg D. et al. (1989), N Engl J Med 320: 915 - 924
2. Witztum J.L. (1994), Lancet 344: 793 - 795
3. Palinski W. et al. (1989), Proc Natl Acad Sci USA 86: 1372 - 1376
4. Esterbauer H. et al. (1991), Free Radical Biol & Med 13: 341 - 390
5. Fogelman A. et al. (1980), Proc Natl Acad Sci USA 77: 2214 - 2218
6. Goldstein J.L. et al (1977), Proc Nat Acad Sci USA 76: 33 - 37
7. Parums D. V. et al. (1990), Arch Pathol Lab Med 114: 383 - 387
8. Salonen J.T. et al. (1992), Lancet 339: 883 - 887
9. Ware D. et al. (1994), Lancet 343: 645 - 646
10. Vaarala O. et al. (1993), Lancet 341: 923 -925
11. Lipoprotein Oxidation and Atherosclerosis (1994), Sept. 17 - 18,Pavia, Italy, G. Finardi, G. Bellomo, E. Maggi
12. Schuhmacher M. et al (1994) Free Red Biol Med
13. Esterbauer H. et al (1989), Free Rad Res Comm 6: 67 - 75
14. Tatzber F. et al. (1991), Atherosclerosis 89: 203 – 208

11. Incubation scheme

	Blank	Standard/Controls	Sample
Buffer	200 µl	200 µl	200 µl
Standards/Controls		20 µl	
Samples			20 µl

incubate the microwell strips covered with plastic film for 90 min. at 37 °C in an incubator
discard liquid and wash the wells 4 times with 300 µl washing solution

Conjugate	100 µl	100 µl	100 µl
-----------	--------	--------	--------

incubate the microwell strips covered with plastic film for 30 min. at room temperature
discard liquid and wash the wells 4 times with 300 µl washing solution

Substrate	100 µl	100 µl	100 µl
-----------	--------	--------	--------

incubate for 15 min. at room temperature in the dark

Stop solution	50 µl	50 µl	50 µl
---------------	-------	-------	-------

read results with an ELISA microwell reader at 450 nm and
620 or 690 nm as a reference

* - prediluted (1 + 4)



DIAGNOSTIC AUTOMATION, INC.

23961 Craftsman Road, Suite E/F, Calabasas, CA 91302

Tel: (818) 591-3030 Fax: (818) 591-8383

ISO 13485-2003

Revision Date: 4/10/06