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IVD



See external label



2°C-8°C



Σ=96 tests

REF

#1431

Ribosomal P IgA, G, M ELISA

Catalog # 1431

INTENDED USE

The Diagnostic Automation, Inc. (DAI) Ribosomal P IgG,A,M Enzyme-Linked Immunosorbent Assay (ELISA) is intended for the detection and semi-quantitative determination of IgG,A,M antibodies to Ribosomal P in human sera. The assay is to be used to detect antibodies in a single serum specimen. The results of the assay are to be used as an aid in the diagnosis of Systemic Lupus Erythematosus (SLE). For *in vitro* diagnostic use.

SUMMARY

Systemic autoimmune disease is characterized by the presence of circulating auto-antibodies directed to a wide variety of cellular antigens (1,2,3). Systemic lupus erythematosus (SLE), commonly referred to as Lupus is the best known of these diseases. Other possible connective tissue diseases include mixed connective tissue disease (MCTD), Sjogren syndrome, scleroderma, and polymyositis/dermatomyositis. The majority can be diagnosed by clinical presentation and their antibody profiles to the various antigens involved, which include dsDNA, SM, RNP, Ro, La, Scl-70, Jo-1 and Histones (1,2,3). Therefore, immunoassays for autoantibodies are useful for diagnostic and prognostic evaluations of autoimmune disease (1,2,3).

PRINCIPLE

The DAI Ribosomal P test is an Enzyme-Linked Immunosorbent Assay to detect IgG,A,M antibodies to Ribosomal P antigens. Purified Ribosomal P antigens are attached to a solid phase microassay well. Diluted test sera are added to each well. If the antibodies are present that recognize the antigen, antigen-antibody complexes are formed. After incubation, the wells are washed to remove unbound antibody. An enzyme labeled anti-human IgG,A,M is added to each well. If antibody is present, the conjugate will bind to the antigen-antibody complexes. After incubation, the wells are washed to remove unbound conjugate. A substrate solution is added to each well. If enzyme is present, the substrate will undergo a color change. After an incubation period, the reaction is stopped and the color intensity is measured photometrically, producing an indirect measurement of specific antibody in the patient specimen.

MATERIALS SUPPLIED

1. Ribosomal P antigen coated microassay plate: 96 wells, provided with a strip holder and stored in a foil bag with desiccant/humidity indicator. (96T: one plate)

2. Serum Diluent: ready to use. Contains proclin (0.1%) as a preservative, pH 7.5 ± 0.2 . (96T: one bottle, 30 mL)
3. Calibrator: human serum. Sodium Azide (0.1%) and penstrep (0.01%) added as preservatives, with kit specific factor printed on vial label. (96T: one vial, 0.4 mL)
4. High Positive Control: human serum. Sodium Azide (0.1%) and penstrep (0.01%) added as preservatives, with established range printed on vial label. (96T: one vial, 0.4 mL)
5. Low Positive Control: human serum. Sodium Azide (0.1%) and penstrep (0.01%) added as preservative, with established range printed on vial label. (96T: one vial, 0.4 mL)
6. Negative Control: human serum. Sodium Azide (0.1%) and penstrep (0.01%) added as preservatives, with established ranges printed on vial label. (96T: one vial, 0.4 mL)
7. Horseradish-peroxidase (HRP) Conjugate: ready to use. Goat anti-human IgG,A,M, containing proclin (0.1%) as a preservative. (96T: one bottle, 16 mL)
8. Chromogen/Substrate Solution: Tetramethylbenzidine (TMB), ready to use. (96T: one bottle, 15 mL)
9. Wash Buffer (20X concentrate): Contains TBS, Tween and proclin (0.1%) as a preservative. (96T: one bottle, 50 mL)
10. Stop Solution: Contains a H_2SO_4 solution, ready to use. (96T: one bottle 15 mL)

PRECAUTIONS

1. Each donor unit used in the preparation of the Calibrator and Controls was tested by an FDA approved method for the presence of the antibody to HIV-1 as well as for hepatitis B surface antigen and found to be negative. Because no test method can offer complete assurance that human immunodeficiency virus (HIV-1), hepatitis B virus, or other infectious agents are absent, these specimen/reagents as well as patient samples, should be handled at the Biosafety Level 2 as recommended for any potentially infectious human serum or blood specimen in the Centers for Disease Control/National Institute of Health manual "Biosafety in Microbiology and Biomedical Laboratories," 1993 (4).
2. Certain reagents in this kit contain sodium azide for use as a preservative. Azides may react with lead and copper plumbing to form explosive azide compounds. When disposing of reagents, flush with copious quantities of water to minimize azide build up.
3. Reagents contain preservatives which may be toxic if ingested.
4. Do not pipette by mouth. Avoid contact of reagents and patient specimens with skin or mucous membranes.
5. Do not allow the stop solution to contact skin or eyes. If contact occurs, immediately flush with copious quantities of water.
6. Avoid splashing or generation of aerosols.
7. Do not use heat inactivated sera.
8. Do not mix or interchange reagents between lots of kits or from other manufacturer.
9. Do not dilute or adulterate kit reagents.
10. Do not cross contaminate reagents or specimens.
11. Do not use TMB Substrate solution if it has begun to turn blue.
12. Reusable glassware must be washed out and thoroughly rinsed free of all detergents.
13. Do not vary reagent and incubation temperatures above or below room temperature (21° - 25° C).
14. Washing is important. Improperly washed wells will give erroneous results. Do not allow the well to dry out between incubations.
15. This product is for IN VITRO DIAGNOSTIC USE only.

MATERIAL REQUIRED BUT NOT PROVIDED

1. Wash bottle, automated or semi-automated microwell plate washing system.
2. Micropipettes, including multichannel, capable of accurately delivering 10 μ L-200 μ L volumes (less than 3% CV).
3. One liter graduated cylinder.
4. Paper towels.
5. Test tubes for serum dilutions.
6. Reagent reservoirs for multichannel pipettes.
7. Pipette tips.
8. Distilled or deionized water, CAP Type 1 or equivalent.
9. Timer capable of measuring to an accuracy of ± 1 second.
10. Disposal basins and 0.5% sodium hypochlorite (50 mL bleach in 950 mL H₂O).
11. Single or dual wavelength microplate reader with 450 nm filter. If dual wavelength is used, set the reference filter to 600-650 nm. Read the Operator's Manual or contact the instrument manufacturer to establish linearity performance specifications of the reader.

STORAGE AND SHELF LIFE OF REAGENTS

1. All kit components that are stored at their recommended storage conditions are stable until the expiration date on their label. Do not use past their expiration date.
2. Antigen coated wells. Unused strips should be immediately resealed in the foil bags with desiccant/humidity indicator and returned to storage at 2°- 8° C. If the bag is resealed with tape the wells are stable for 30 days. If the bag is resealed with a heat sealer the wells are stable until their expiration.
3. All other reagents are stored at 2°- 8° C in their original containers.
4. Store 1X (diluted) Wash Buffer at room temperature (21°- 25 ° C) for up to 5 days, or 1 week between 2° - 8° C.

SPECIMEN COLLECTION

1. Aseptically collect blood samples by venipuncture and prepare serum using accepted technique (5).
2. Serum containing visible particulate matter can be spun down utilizing slow speed centrifugation.
3. Sera may be stored up to five days at 2°- 8° C. If a further delay in testing is needed, store frozen at -20° to -70° C in a non-defrosting freezer. Avoid multiple freeze/thaw of patient samples.
4. Avoid using hemolyzed, lipemic, or bacterially contaminated sera.
5. Do not heat inactivate sera.

PREPARATION OF REAGENTS

1. All reagents must be removed from refrigeration and allowed to come to room temperature (21°- 25° C) before use. Return all reagents to refrigerator promptly after use.
2. All samples and controls should be vortexed before use.
3. Dilute 50 mL of the 20X Wash buffer to 1L with distilled and/or deionized H₂O. Mix well.

GENERAL PROCEDURE

1. Determine the number of patients to be assayed. For each assay, the Calibrator should be run in triplicate. Also, the High Positive Control, Low Positive Control, Negative Control, and a reagent blank (RB) should be run on each assay. Check software and reader requirements for the correct Calibrator/Control configurations.

Example Configuration:

1A	RB	2A	Patient #2
1B	NC	2B	Patient #3
1C	Cal	2C	Patient #4
1D	Cal	2D	Patient #5
1E	Cal	2E	Patient #6
1F	HPC	2F	Patient #7
1G	LPC	2G	Patient #8
1H	Patient #1	2H	Patient #9

2. For each serum assayed, prepare a 1:21 serum dilution. Add 10 μ L of each serum sample to 200 μ L of Serum Diluent. Mix well.
3. Remove the number of wells needed from the plate bag and arrange in a strip holder. The remaining strips should be resealed in the plate bag with desiccant/humidity indicator. The bag should be reheat sealed or rolled over and the end taped. If the color of the indicator changes from blue to pink, the strips should not be used.
4. Transfer 100 μ L of the prediluted samples to the reaction wells, using a multichannel pipette. Withdraw and expel each sample at least three times to ensure proper mixing of the sample before transferring to the reaction plate. Use new fresh pipette tips for each sample. Add 100 μ L of Serum Diluent to the reagent blank.
5. Incubate each well of the reaction plate at room temperature (21°- 25° C) for 30 minutes \pm 1 minute.
6. Wash the reaction plate three times with 1X Wash Buffer. Shake all of the liquid out of the wells. With a wash bottle, automated or semi-automated wash system, fill each well with Wash Buffer making sure no air bubbles are trapped in the wells. Shake all of the Wash Buffer out of the wells. Repeat the wash two more times. A total of up to 5 washes may be necessary with automated equipment. After the last wash, shake out the Wash Buffer and remove residual Wash Buffer by tapping the plate firmly on a paper towel. The Wash Buffer can be collected in a basin and treated with 0.5% sodium hypochlorite (bleach) at the end of the day.
7. Add 100 μ L of the Conjugate to each well of the reaction plate, including reagent blank.
8. Incubate each well of the reaction plate at room temperature (21°- 25° C) for 30 minutes \pm 1 minute.
9. Repeat wash as described in Step 6.
10. Add 100 μ L of the Chromogen/Substrate Solution to each well of the reaction plate, including reagent blank.
11. Incubate each well of the reaction plate at room temperature (21°- 25° C) for 15 minutes \pm 1 minute.
12. Add 100 μ L of the Stop Solution to each well at the same rate as the TMB Substrate was added. Positive samples will turn from blue to yellow. Tap plate to ensure mixing. Wait a minimum of 5 minutes and read.

13. Read the plate using a spectrophotometer at a wavelength of 450 nm. If dual wavelength is used, set the reference filter to 600-650 nm. Measure each optical density (OD) against the reagent blank. The plate should be read within 30 minutes of assay completion.

QUALITY CONTROL

1. Calibrator and Controls must be run with each test run.
2. Reagent Blank must be < 0.15 O.D. at 450 nm.
3. The mean O.D. Value for the Calibrator should be ≥ 0.30 at 450 nm.
4. The Index Values for the High, Low, and Negative Controls should be in their respective ranges printed on the vials. If the control values are not within their respective ranges, the test should be considered invalid and the test should be repeated.
5. If above criteria are not met on repeat, contact DAI Technical Service.

CALCULATION OF RESULTS

1. Calibrator Value - Calculate the mean value for the Calibrator from the three Calibrator determinations. If any of the three Calibrator values differ by more than 15% from the mean, discard that value and calculate the mean of the two remaining values
2. Correction Factor - To account for day-to-day fluctuations in assay activity due to room temperature and timing, a Correction Factor is determined by DAI for each lot of kits. The Correction Factor is printed on the Calibrator vial.
3. Cutoff O.D. Value - The Cutoff O.D. Value for each assay is determined by multiplying the Correction Factor by the mean Calibrator Value determined in step 1.
4. Index Value - Calculate an Index Value for each specimen by dividing the specimen O.D. Value by the Cutoff O.D. determined in step 3.

Example : O.D.s obtained for Calibrator = 0.38, 0.40, 0.42
 Mean O.D. for Calibrator = 0.40
 O.D. obtained for patient sera = 0.60
 Correction Factor = 0.50
 Cutoff Value = 0.50 x 0.40 = 0.20
 Index Value = 0.60/0.20 = 3.00

INTERPRETATION OF RESULTS

<u>Index Value</u>	<u>Interpretation</u>
≤ 0.90	Negative
0.91 - 1.09	Equivocal
≥ 1.10	Positive

Specimens with Index Values in the equivocal range should be retested. If still equivocal, retest by an alternate method or test a new sample.

LIMITATIONS

1. The result of the assay should not be interpreted as being diagnostic. The results should only be used as an aid to diagnosis. The results should be interpreted in conjunction with the clinical evaluation of the patient.
2. The assay should be used only with serum. Icteric, lipemic, hemolyzed and heat inactivated serum should be avoided.

3. Index Values of > 10.00 should be reported as greater than 10.
4. Specimens with Index Values in the equivocal range should be retested. If still equivocal, retest by an alternate method or test a new sample.

EXPECTED VALUES

1. To determine the prevalence of Ribosomal P antibody in lupus patients, 451 sera from patients from a lupus cohort were tested on the DAI Ribosomal P IgG,A,M ELISA kit. Forty five sera were found to be positive for a prevalence rate of 9.98%. The data indicate that the prevalence of Ribosomal P antibody in a lupus population is similar to that found in the literature (12-20%).
2. Antibodies to Ribosomal P are rare in the normal population.

PERFORMANCE CHARACTERISTICS SENSITIVITY AND SPECIFICITY

The DAI Ribosomal P IgG,A,M ELISA test results were compared to results obtained by Ouchterlony analysis of serum from clinically defined Lupus (n=46) and normals (n=137). Table 1 summarizes the data.

Table 1
Sensitivity and Specificity of the DAI Ribosomal P IgG,A,M ELISA Kit

		DAI Ribosomal P ELISA Kit			
		Positive ≥ 1.10	Equivocal 0.91-1.09	Negative ≤ 0.90	Total
Ouchterlony	Positive	46	0	0	46
	Negative	1	0	136	137
	Total	47	0	136	183
Relative Sensitivity	= 46/46	= 100%	95% Confidence Interval = 93.5% - 100%		
Relative Specificity	= 136/137	= 99.3 %	95% Confidence Interval = 97.8% - 100%		
Relative Agreement	= 182/183	= 99.5 %	95% Confidence Interval = 98.4% - 100%		

The 95% confidence interval for relative sensitivity was calculated assuming one false negative.

PRECISION

The precision of the DAI Ribosomal P IgG,A,M ELISA kit was determined by testing six different sera eight times each on three different assays. Table 2 summarizes the data. With proper technique, the user should obtain C.V.'s of less than 15%.

Table 2
Ribosomal P Precision Data

<u>Serum #</u>	<u>Assay 1 (n=8)</u>			<u>Assay 2 (n=8)</u>			<u>Assay 3 (n=8)</u>			<u>Inter Assay (n=24)</u>		
	<u>X</u>	<u>S.D.</u>	<u>C.V.</u>	<u>X</u>	<u>S.D.</u>	<u>C.V.</u>	<u>X</u>	<u>S.D.</u>	<u>C.V.</u>	<u>X</u>	<u>S.D.</u>	<u>C.V.</u>
1	1.57	0.117	7.4%	1.62	0.113	7.0%	1.50	0.134	8.9%	1.57	0.127	8.1%
2	1.68	0.176	10.5%	1.60	0.095	5.9%	1.66	0.148	8.9%	1.65	0.143	8.7%
3	2.98	0.113	3.8%	2.83	0.128	4.5%	2.74	0.127	4.6%	2.85	0.155	5.5%
4	2.89	0.115	4.09%	2.95	0.135	4.6%	2.94	0.068	2.3%	2.92	0.109	3.7%
5	0.29	0.027	9.29%	0.18	0.035	19.6%	0.20	0.039	19.6%	0.22	0.056	25.6%
6	0.09	0.042	46.3%	0.08	0.016	20.5%	0.08	0.037	46.5%	0.10	0.048	48.0%

X = Mean Ribosomal P Value

S.D. = Standard Deviation

C.V. = Coefficient of Variation

LINEARITY

The DAI Ribosomal P IgG,A,M ELISA Index Values were determined for serial twofold dilutions of five positive sera. The Index Values were compared to \log_2 of dilution by standard linear regression. The data in Table 3 indicates that the assay is semi-quantitative.

Table 3
Linearity

<u>Serum #</u>	<u>Neat</u>	<u>1:2</u>	<u>1:4</u>	<u>1:8</u>	<u>1:16</u>	<u>1:32</u>	<u>1:64</u>	<u>1:128</u>	<u>r</u>
1	2.70	2.52	2.24	1.93	1.57	1.33	0.98		0.997
2	2.53	2.29	1.99	1.57	1.20	0.86			0.997
3	2.89	2.71	2.47	2.23	1.93	1.57	1.20	0.90	0.994
4	3.68	3.18	2.59	1.94	1.59	1.00	0.68		0.997
5	2.09	1.38	0.94						0.991

CROSS REACTIVITY

Sera containing high levels of antibodies to potentially cross reactive antigens were assayed on the DAI Ribosomal P IgG,A,M ELISA kit. The data in Table 4 indicate that antibodies to alternate autoimmune antigens do not cross react with the DAI Ribosomal P ELISA kit.

Table 4
Cross Reactivity

<u>Serum #</u>	<u>DAI Index Value</u>	<u>Interpretation</u>	<u>Specificity</u>
1	0.21	-	Ro
2	0.18	-	Ro
3	0.17	-	Ro
4	0.13	-	La
5	0.09	-	La
6	0.10	-	La
7	0.09	-	Scl-70
8	0.18	-	Scl-70
9	0.17	-	Scl-70
10	0.15	-	Jo-1
11	0.18	-	Jo-1
12	0.15	-	Jo-1
13	0.38	-	Sm
14	0.43	-	Sm
15	0.40	-	Sm
16	0.19	-	RNP
17	0.15	-	RNP
18	0.14	-	RNP
19	0.14	-	DNA

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ISO 13485-2003

